

Effect of Cyclohexenonic Long Chain Fatty Alcohols on Neurite Outgrowth. Study on Structure-Activity Relationship

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Abstract: Four series of long chain fatty alcohols bearing a cyclohexenone moiety in addition to a ω-alkanol side chain were synthesized using "Umpolung" reactivity strategy. Their effect on neurite outgrowth was evaluated by means of fetal rat neurons in culture. The length of the ω-hydroxy side chain is a crucial factor for biological activity. © 1998 Elsevier Science Ltd. All rights reserved.

INTRODUCTION

The normal development of the vertebrate nervous system entails the death of 30-70% of the neurons originally generated in most neuronal populations ¹. This naturally occurring cell death is regulated by specific neurotrophic factors for exemple NGF (Nerve Growth Factor), BDNF (Brain Derived Neurotrophic Factor), NT-3 (Neurotrophin-3), NT-4/5 (Neurotrophin-4/5), FGF (Fibroblast Growth Factor)² etc. They are produced in limited amounts by target cells, such as glial cells and promote neuronal survival and differentiation. These factors may potentially lead to a viable therapy to alter the pathogenesis of neurodegenerative diseases. However, these high molecular weight polypeptides are not able to cross several biological barriers, in particular the blood brain barrier, and consequently they can not be active therapeutic agents for the treatment of neurodegenerative diseases.

To overcome these difficulties, it has been suggested that small lipids which are able to mimick the biological activity of these naturally-occurring protein neurotrophic factors, to stimulate their production or to enhance their activity³ would be ideal targets for research.

Recently, several examples of low-molecular weight compounds exhibiting neurotrophic properties have been reported. Some of these act indirectly by inducing the synthesis and the release of NGF. They are derivatives of catecholamines⁴, of benzoquinones⁵, hericenone⁶, fellutamide⁷, dictyophorine⁸ and erinacine⁹. The others act directly on neurons and promote a neuronal differentiation effect by inducing neurite outgrowth. These are molecules such as lactacystin^{10,11}, epolactaene¹² and also extend to sesquiterpene-neoligans¹³, SR 57746A¹⁴, 1,3,3 tricyano-2-amino-1-propene¹⁵ and PS-990¹⁶.

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Our initial research has lead to the demonstration of the neurotrophic activity of n-hexacosanol¹⁷, a C₂₆ primary alcohol and of analogues of perhydroretinol¹⁸. Indeed these substances are able to promote the survival and the differentiation of rat central nervous system neurons in culture^{19,20}. At 10⁻⁷-10⁻⁸M, they induce markedly neurite outgrowth and appearance of multipolarity on treated neurons.

These fatty alcohols contain a large hydrophobic side chain and a small primary hydroxy group. We have postulated the necessary presence of a receptor or a carrier for these molecules mainly because of the specificity of the effect. Indeed in the straight chain series, the C₂₆ hexacosanol is the most potent compound while the C₂₈ octacosanol is only slighty active and the C₂₄ tetracosanol and the C₃₀ triacontanol are inactive. Furthemore, in the perhydroretinol series, compounds containing a C₁₁ side chain are the most active.

In addition to a hydrophobic effect, another interaction is due to the presence of the alcoholic function. At this point we now focused our attention on a series of long chain fatty alcohols bearing a ketone group. This second polar group may act as a second binding site and this new interaction leads to a more potent effect. In this paper, we report the synthesis of 4 series of long chain fatty alcohols bearing a cyclohexenonic function and an ω -alkanol containing 10 to 16 carbon atoms. These compounds are derivatives of cyclohexenone (compounds 1a), of 4-methyl-cyclohexenone (compounds 1b), of 4,4-dimethyl-cyclohexenone (compounds 1c) and of 2-methyl-cyclohexenone (compound 1d) (scheme 1). The preliminary data of their neurotrophic activity showed that compounds 1b and 1d with a C_{14} side chain are the most potent. These molecules are labelled as 1b (n=14) and 1d (n=14).

<u>Scheme 1</u>: The 4 series of cyclohexenonic alcohols differred by the presence of methyl groups in the cyclohexenonic ring.

RESULT AND DISCUSSION

Four series of long chain fatty alcohols 1a to 1d were synthesized by coupling an ω -alkanol side chain to different cyclohexenones. As we wished to introduce the alkanol side chain by means of a nucleophilic displacement it was necessary to "invert" the polarity of the cyclohexenone moiety in the synthetic strategy.

1a to 1d were synthesized in the same manner. The starting materials were respectively 2a,3a,2b and 3b (scheme 2).

A) Synthesis of 1a, 1b, 1c, 1d long chain fatty alcohols

Scheme 2

I R₁ R₂
$$R_3$$
 R_3 R_3 R_3 R_4 R_4 R_5 R_5 R_5 R_5 R_5 R_6 R_6 R_7 R_8 R_9 R_9

Scheme 2 shows the retrosynthetic analysis for the preparation of 1a, 1b, 1c, 1d series. I can be broken up into the sulfone II and the side chain III.

Synthesis of cyclohexenones

2b $R_1=R_2=Me$, $R_3=H$ 3b $R_1=R_2=H$, $R_3=Me$

2a and 2b are commercially available products but 4-methyl-2-cyclohexen-1-one and 2-methyl-2-cyclohexen-1-one have to be synthesized from 3a and 3b respectively (scheme 3,4).

As shown in **scheme 3**, enol silane **4a** was formed using lithium diisopropylamide and trimethylsilyl chloride. The formation of the double bond was achieved using a catalytic amount of Pd(OAc)₂ in DMSO under a dioxygen atmosphere ²¹ to give **5a**. It was be tested at present state as racemics.

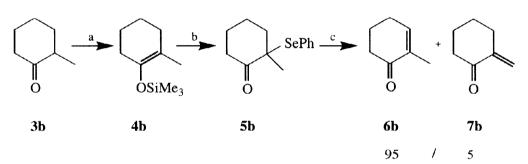
Scheme 3

Reagents: a) 1) LDA, -78°C, 1h, 2) Me₃SiCl, -78°C, 1h (96%); b) Pd(OAc)₂ cat, DMSO, O_2 , r.t., 6h (72%).

However, this latter technique was not appropriate for the preparation of 2-methyl-2-cyclohexen-1-one **6b** which was synthesized as shown in **scheme 4**.

Enol silane **4b** was prepared with disopropylamide magnesium bromide and trimethylsilyl chloride. **4b** was subsequently reacted with phenyl selenium chloride followed by oxidation and elimination of selenium using hydrogen peroxide. **6b** and **7b** were obtained with a ratio of 95/5 (scheme **4**).

Scheme 4



Reagents: a) 1) (iPr)₂NMgBr, Et₂O, r.t., 12h, 2) Me₃SiCl, Et₃N, HMPA, r.t., 3h (85%); b) PhSeCl, THF, -78°C, 10 min (85%); c) H_2O_2 , CH_2Cl_2 , pyridine, 0°C, 1h (98%).

Synthesis of ω-bromoalcohols

11-Bromo-undecanol is commercially available while 10-bromodecanol and 12-bromododecanol were synthesized from the corresponding diols by means of monobromation in a mixture of HBr-cyclohexane. The bromoalcohols were obtained in over 80% yield and the dibromoalkane in 10% yield (scheme 5).

Scheme 5

HO OH
$$\frac{\text{HBr } 47\%}{\text{cyclohexane}}$$
 Br OH + Br Br $\frac{\text{Br}}{\text{n}}$ Br $\frac{\text{Br}}{\text{n}}$ $\frac{\text{Br}}{\text{n}}$

The C₁₃,C₁₄,C₁₅ and C₁₆ bromoalcohols were synthesized in two steps from the corresponding diacids. The first step consisted of a reduction of the diacids to the diols with lithium aluminium hydride and the second step was the monobromination. (scheme 6).

Scheme 6

HOOC COOH LIAIH4 HO OH Cyclohexane

11 12
$$n = 13 (93\%)$$
 $n = 14 (91\%)$ $n = 15 (97\%)$ $n = 16 (84\%)$ $n = 16 (80\%)$

Coupling between cyclohexenones and the side chain

As shown in **scheme 7**, to reverse the polarity of the differents cyclohexenones we used a phenyl sulfonyl group which allowed the formation and the stabilisation of a carbanion in the α position.

In an aqueous medium, 14 reacted with benzenesulfinic acid sodium salt to give sulfone 15^{22} . The carbonyl function of the latter was protected by the formation of the ketal 16. The carbanion was generated using n-butyllithium in the presence of HMPA and reacted with the unprotected ω -bromoalcohols. 17 was obtained in 84-98%.yield.

The desulfonylation-deketalization was accomplished in one step by treating 17 with p-TsOH (5% mol/equiv) in chloroform/acetone 5/1 at 50°C ²³.18 was obtained in 58-90%.yield

Scheme 7

$$R_1$$
 R_2
 R_3
 R_3
 R_3
 R_4
 R_5
 R_5
 R_5
 R_5
 R_5
 R_5
 R_7
 R_7

Reagents : a) PhSO₂Na, HCl, H₂O, r.t., 24h; b) HO-(CH₂)₂-OH, pTsOH, benzene, reflux, 4h; c) 1) nBuLi, THF, HMPA, -78°C to r.t., 1h, 2) Br-(CH₂)_n-OH, THF, -78°C to -20°C, 2h; d) CHCl₃-acetone (5-1), pTsOH, 50°C, 24h.

B) Effect of different cyclohexenonic fatty alcohols on primary neuronal cultures

To investigate the differentiation effect of cyclohexenonic fatty alcohols, we undertook studies of the two most typical criteria of neuronal differentiation. We quantified the enhancement of neurite outgrowth and analysed the modification of morphology of treated neurons. It is well known that a high percentage of long neurites and of neuron multipolarity is associated with a high degree of differentiation.

Fatty alcohols **1a** which do not contain methyl group on cyclohexenonic ring do not enhance neurite outgrowth. Moreover, they are toxic at 10⁻⁵M.

Table 1, shows the comparative effect of cyclohexenonic fatty alcohols that we have synthesized. Neurite outgrowth is induced by fatty alcohols bearing a side chain containing at least 12 carbon atoms. Compounds 1b and 1d with a methyl group at C-4 and C-2 are the most potent. And among these, the most active carry a C₁₄ side chain. With a slightly longer C₁₅ side chain, the activity disappears completely and moreover is replaced by a neurotoxicity

	COMPOUNDS	ACTIVITY
	CONTROL	0
	bFGF	++
1a	(n = 11)	
	(n = 12)	
	(n = 13)	-
	(n = 14)	-
1b	(n = 10)	+
1	(n = 11)	+
	(n=12)	+++
l	(n = 13)	++++
	$(\mathbf{n}=14)$	++++
	(n=15)	0
	(n = 16)	0
1c	(n = 10)	0
	(n = 11)	++
	$(\mathbf{n}=12)$	+++
	(n = 13)	+++
	(n = 14)	+++
1d	(n = 10)	+
1	(n = 11)	0
1	(n=12)	++
1	(n=13)	+++
1	$(\mathbf{n}=14)$	++++
1	(n=15)	
L .	(n = 16)	

Table 1

Effect of different ketonic long chain fatty alcohols on neurite outgrowth: At least two independent experiments were performed for each treatment. The compound dissolved in EtOH was added to the culture medium after seeding at 10⁻⁸ M. Plus signs indicate a significant difference in the percentage of cells exhibiting a bipolar morphology as well as neurites two times longer than in control. An additional plus sign indicates that the differentiated cells exhibited a predominantly multipolarity as well as neurites more than two times longer than the control. Minus signs indicate a toxic effect with decrease of neuron population.

Fatty alcohols 1b(n=14) and 1d(n=14) enhance neurite outgrowth and induce appearance of multipolarity at $10^{-8}M$.

To investigate the potency of neurite outgrowth in treated neurons, we classified the neurons according to the total length of their neurites. Five types of neurons were taken under consideration. As shown in **Figure 1**, in the control culture, 90% of the neurons bear a neurite shorter than 208µm (types 1 and 2). In culture treated by **1b** (n=14) and **1d** (n=14), over 24% of neurites are longer than 208µm (types 3, 4 and 5). The effect of bFGF is less potent than that of fatty alcohols.

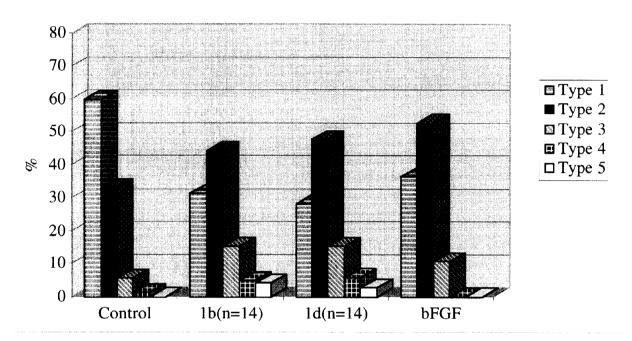


Figure 1: Neurite outgrowth induced by cyclohexenonic long chain fatty alcohols: Neurons are classified according to the total lenght of their neurite: type 1 = less than 104 μ m; type 2: between 104 and 208 μ m; type 3 = between 208 and 311 μ m; type 4 = between 311 and 415 μ m and type 5 over 415 μ m. The figure shows the % of these different types of neurons after treatment with the 2 most potent cyclohexenonic derivatives: 1b (n = 14) and 1d (n = 14) in comparison with control culture and with the effect of bFGF in the same conditions. The total of neurons of all types represents 100%. The length of neurites was measured by NIH Image.

To study the ability of fatty alcohols to promote the appearance of multipolarity, we quantified neurons with a special morphology for example: neurons bearing neurites longer than 141µm, neurons with a long neurite and many collaterals, as well as stellate shaped multipolar neurons. As shown in **Figure 2**, only 10% of control neurons belong to one of these 3 groups while 50% of fatty alcohol treated neurons are endowed with special morphology. Once again, the effect of bFGF is weaker than that of fatty alcohols.

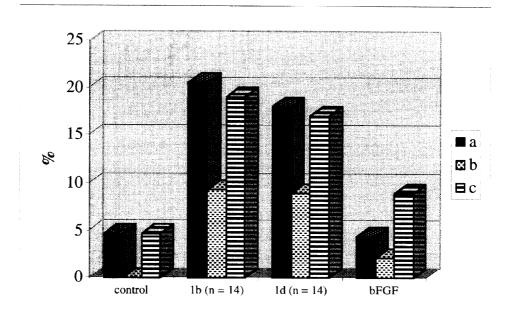


Figure 2:% of special neurons after treatment with cyclohexenonic long chain fatty alcohols. Neurons are divided into 3 types according to their special morphology a) neurons with at least one long neurite > 141 μ m, b) neurons with one long neurite > 141 μ m and bearing >2 collaterals, c) stellate-shaped multipolar neurons. Ketonic compounds 1b (n = 14) and 1d (n = 14) induce more of these special types of neurons which represent a high degree of differentiation.

Our data shed light on a structural requirement which is crucial for the neurotrophic activity of long chain fatty alcohols. The presence of a methyl group in the cyclohexenonic ring plays an important role. Indeed compounds 1a which contain no methyl group are inactive.

The crucial role played by the length of the side chain appears in every series of fatty alcohols that we have investigated. In the straight chain series, the C_{26} hexacosanol is the most active while the C_{28} octacosanol is only slightly active and the C_{30} triacontanol and the C_{24} tetracosanol are totally inactive. In the perhydroretinol series, compounds bearing a C_{11} side chain are by far the most neurotrophic. And in the cyclohexenonic series , which is described in the present publication, an ω -tetradecanol side chain induces the most potent differentiation effect in comparison to a longer or a shorter side chain.

EXPERIMENTAL SECTION

Tetrahydrofuran (THF) was distilled from sodium/benzophenone under argon prior to use. Dichloromethane, methanol and benzene were distilled from calcium hydride. Pyridine was distilled from potassium hydroxide. Hexamethylphosphoramide (HMPA) was distilled from calcium hydride and stored over 3Å molecular sieves under argon. All reactions involving moisture sensitive reactants were executed under an argon atmosphere using oven dried and/or flame dried glassware.

reactants were executed under an argon atmosphere using oven dried and/or flame dried glassware.

¹H NMR spectra were recorded on Bruker SY 200 (200 MHz) and AM 400 (400 MHz) spectrometers as solutions in deuterochloroform (CDCl3). Chemical shifts are expressed in parts per million (ppm, δ) downfield from tetramethylsilane (TMS) and are referenced to CHCl3 (7.26 ppm) as internal standard. Splitting patterns are designated as s, singlet; d, doublet; t, triplet; q, quartet; qn, quintuplet; m, multiplet; br, broad. Coupling constants are given in hertz (Hz). ¹³C NMR spectra were recorded on Bruker SY 200 (50 MHz) and AM 400 (75 MHz) spectrometers as solutions in CDCl3. Chemical shifts are reported in parts per million (ppm, δ) downfield from TMS and are referenced to the center line of CDCl3 (77.0 ppm) as internal standard (*, c* interchangable assignments). The attribution of the different carbons (C, CH, CH2, CH3) was determined by ¹³C to ¹H polarisation transfer (DEPT). IR spectra were recorded on sodium chloride using Perkin-Elmer 881 FT-IR spectrometer and are reported in wave numbers (cm⁻¹). UV spectra were obtained in acetonitrile solution using a Kontron-Uvikon 810 UV-Vis spectrometer. Mass spectra (MS) were measured on a TRIO 2000 apparatus by direct introduction (an ionisation potential of 70 eV was used, m/z relative intensities (in %) are noted in round brackets) or coupled to a GC DB5 column (J.W). GC chromatograms were obtained by dissolving the sample in ethyl acetate, using a Carlo-Erba apparatus and a SE 30 column. Microanalyses were performed by the Service Central de Microanalyse du CNRS (Strasbourg). Routine monitoring of reactions were performed using 60 F₂₅₄ silica gel TLC plates (Merck), which were dipped in a solution of vanillin (1 g) in EtOH/H2SO4 (95/5) and heated on a hot plate. Flash chromatography was conducted using 60 F₂₅₄ silica gel (Merck) with the indicated solvent.

4-Methyl-1-(trimethylsilyloxy)-1-cyclohexene 4a

Synthesis of lithium diisopropylamide (LDA)

To a solution of N,N-diisopropylamine (7 ml, 49.6 mmol, 1.5 eq.) in 20 ml of THF was added dropwise a solution of n-BuLi (1.4M, 35.4 ml, 49.6 mmol, 1.5 eq) at -78°C. The solution was stirred at 0°C for 30 min. Synthesis of 4a

To a solution of 4-methylcyclohexan-1-one **3a** (4 ml, 33 mmol, 1 eq.) in 10 ml of THF was added the preceding solution at -78°C. The mixture was stirred at -78°C for 1 h, and trimethylsilyl chloride (6.5 ml, 1.5 eq) was added. After stirring at room temperature for 1 h the solution was poured into a solution of NaHCO3 and extracted with ether. The organic layer was dried over MgSO4 and concentrated. The residue was purified by distillation under vacuo to obtain a liquid **4a** (5.83 g, 96%). **TLC**: (hexane-AcOEt: 8-2) Rf = 0.8; **GC**: 30°C (7 min) --> 150°C (15°C/min) --> 280°C (20°C/min) 11.1 min, 96%; ¹H NMR (200 MHz), δ : 0.17 (s, 9H, Si-(CH₃)₃); 0.94 (d, J = 6.2 Hz, 3H, H-7); 1.2-1.43 (m, 1H, H-4); 1.57-1.76 (m, 4H, H-3,6); 1.88-2.14 (m. 2H, H-5); 4.8-4.83 (m, 1H, H-2); ¹³C NMR (50 MHz), δ : 0.3 (Si-(CH₃)₃); 21.2 (C-7); 28.3 (C-4); 29.6 (C-5); 31.3 (C-6); 32.3 (C-3); 103.5 (C-2); 150.1 (C-1); **IR** (NaCl): 3052, 3021 (w, =C-H); 2954, 2926 (s, C-H); 1670 (s, C=C); 1457 (m); 1371 (m); 1252 (s); 1190 (s); 1046 (m); 892 (s); 844 (s).

4-Methyl-2-cyclohexen-1-one 5a

To a solution of **4a** (3.53 g, 19.2 mmol, 1 eq.) in dry DMSO (70 ml) was added palladium (II) acetate (432 mg, 0.1 eq.). The mixture was stirred under oxygen for 6 h, then water was added at 0°C, the solution was filtrated on celite^R and extrated with ether. The organic layer was concentrated in vacuo and the residue dissolved in a mixture of hexane-water. The solution was extracted with hexane, washed with brine, dried with MgSO₄ and concentrated in vacuo to give a liquid (1.52 g, 72%). **TLC**: (hexane-AcOEt: 8-2) Rf = 0.35; **GC**: 30°C (7 min) --> 150°C (15°C/min) --> 280°C (20°C/min) 5 min; ¹H NMR (200 MHz), δ : 1.15 (d, J = 7.1 Hz, 3H, H-7); 1.56-1.76 (m, 1H, H-5a); 2.1 (dq, Jgem = 13.3 Hz, 3 J = 4.9 Hz, 1H, H-5e); 2.26-2.48 (m, 2H, H-6); 2.49-2.62 (m, 1H, H-4); 5.94 (dd, 3 J = 10.1 Hz, 4 J = 2.5 Hz, 1H, H-2); 6.79 (ddd, 3 J = 10.1 Hz, 3 J = 2.7, 4 J = 1.5 Hz, 1H, H-3); 13°C NMR (50 MHz), δ : 20.1 (C-7); 29.6 (C-5); 30.9 (C-4); 36.8(C-6); 128.4 (C-2); 156.2 (C-3); 199.7 (C-1); IR (NaCl): 3025 (w, =C-H); 2958, 2932 (s, C-H); 1683 (s, C=O); 1617 (w, C=C); 1458 (m); 1391, 1375 (m); 1251 (s); 1094, 1053, 1016 (m); 828 (s); 750 (m).

2-Methyl-1-(trimethylsilyloxy)-1-cyclohexene 4b

Preparation of BMDA (Bromide magnesium diisopropyl amide).

To a solution of diisopropylamine (4.4 ml, 30.9 mmol, 1.25 eq.) in ether (160 ml) was added dropwise a solution of methylmagnesium bromide (3M in ether, 9.8 ml, 29.6 mmol, 1.2 eq.) at room temperature. The suspension was stirred at this temperature for 12 h.

Synthesis of 4b

To the previous solution was added slowly 2-methylcyclohexan-1-one **3b** (3ml, 24.7 mmol, 1eq.) in ether (60ml). After 15 min. trimethylsilyl chloride (9.4ml, 74.1 mmol, 3 eq.), distilled trimethylamine (11.2ml, 80.4 mmol, 3.25 eq) and distilled hexamethylphosphoramide (2.2ml, 0.5 eq) were added. After stirring at room temperature for 3 h, the mixture was poured cautiously into an aqueous solution of NaHCO₃ (2N) at 0°C, then extracted with ether (3 times), washed with brine, dried with MgSO₄, filtered and concentrated in vacuo. The residue was purified by distillation under reduced pressure (0.5 mm Hg) to give a colorless liquid (3.88 g, 85%). TLC: (hexane-AcOEt: 8-2) Rf = 0.7; GC: 30°C (7 min) --> 150°C (15°C/min) --> 280°C (20°C/min) 11.3 min, 97%; ¹H NMR (200 MHz), δ : 0.16 (s, 9H, Si-(CH₃)₃); 1.44-1.70 (m, 4H, H-4, 5); 1.54 (s br, 3H, H-7); 1.88-2.06 (m, 4H, H-3, 6); ¹³C NMR (50 MHz), δ : 0.7 (Si-(CH₃)₃); 16.3 (C-7); 23 (C-4); 23.8 (C-5); 30.1 (C-6*); 30.3 (C-3*); 111.7 (C-2); 142.9 (C-1); IR (NaCl): 2933 (s, C-H); 1716 (s, C=C); 1450, 1377, 1350 (m); 1252, 1184, 1171 (s); 919, 886 (m); 844 (s); MS (EI): 184 (M⁺, 49); 169 (M-CH₃, 91); 155 (21); 141 (50); 75 (82); 73 (SiMe₃, 100); 45 (25).

2-Methyl-2-(phenylselenyl)-cyclohexan-1-one 5b

To a solution of ether **4b** (3.05 g, 16.5 mmol, 1 eq.) in THF (20 ml) was added dropwise PhSeCl (Aldrich, 3.17 g, 1eq) in THF (40 ml) at -78°C. After stirring for 10 min, the mixture was quenched with water and extracted with ether (3 times). The combined organic layers were washed with a saturated aqueous solution of NaHCO3, then with brine, dried with MgSO4, filtered and concentrated in vacuo. The residue was purified by chromatography over silica gel (hexane-AcOEt: 98-2 as eluent) to give an oil (3.7 g, 85%). TLC: (hexane-AcOEt: 9-1) Rf = 0.3; GC: 30°C (7 min) --> 150°C (15°C/min) --> 280°C (20°C/min) 17.2 min. >99%; 1 H NMR (200 MHz), δ : 1.36 (s, 3H, H-7); 1.62-1.84 (m, 2H, H-4); 1.87-1.98 (m, 1H, H-6e); 2-2.2 (m, 2H, H-5); 2.24-2.38 (m, 2H, H-3); 3.42 (ddd, J_{gem} = 14.9 Hz, 3 J = 13.8 Hz, 3 J = 6.2 Hz, 1H, H-6a); 7.23-7.38 (m, 3H, H ar.-3', 4'); 7.43-7.48 (m, 2H, H ar.-2'); 13 C NMR (50 MHz), δ : 22.3 (C-4); 24.8 (C-7); 27.0 (C-5); 37.1 (C-3); 40.7 (C-6); 55.2 (C-2); 126.6 (C ar.-1'); 128.8 (C ar.-2'*); 129.1 (C ar.-4'); 137.4 (C ar.-3'*); 207.5 (C-1).

2-Methyl-2-cyclohexen-1-one 6b

Hydrogen peroxide H₂O₂ (30 % in water, 14.7 ml, 10 eq.) was added dropwise at 0°C to a solution of ketone **5b** in pyridine (2.1 ml, 2 eq) and CH₂Cl₂ (50 ml). After 1h, the solution was refluxed for 5 min, then HCl 2N was added and the mixture was extracted with ether (3 times). The organic layer was dried with MgSO₄, filtered and evaporated. The residue was purified by distillation under reduced pressure to give a colorless liquid (1.4 g, 98%).

TLC: (hexane-AcOEt: 8-2) Rf = 0.35; GC: 30°C (7 min) --> 150°C (15°C/min) --> 280°C (20°C/min) Retention time: α, β-unsaturated ketone **6b**: 6.7min (5%); cyclohexenone **5b**: 7.3min (95%); 1 H NMR (200 MHz), δ: 1.76 (dd, 4 J = 3.4 Hz, 5 J = 2 Hz, 3H, H-7); 1.97 (q, J = 6.4 Hz, 2H, H-5); 2.26-2.36 (m, 2H, H-4); 2.42 (t, J = 6.4 Hz, 2H, H-6); 6.74 (m, 1H, H-3); 13 C NMR (50 MHz), δ: 15.7 (C-7); 23.1 (C-5); 25.8 (C-4); 38.1 (C-6); 135.4 (C-2); 145.3 (C-3); 199.8 (C-1).

10-Bromo-decan-1-ol 9

To a solution of 1,10-decandiol (10 g, 57 mmol) in cyclohexane (150 ml) was added HBr (47% in water, 150 ml). After stirring at reflux for 6h, the solution was extracted with hexane (3 times), washed with saturated NaHCO3 (4 times) and brinc, dried with MgSO4 and concentrated *in vacuo*. The residue was purified by chromatography on silica-gel, eluting with hexane-AcOEt (95-5 to 70-30) to give a colorless oil (12 g, 88%). TLC: (hexane-AcOEt: 6-4) Rf = 0.4; GC: 40-280°C (15°/min) 8.5 min, >99%; 1 H NMR (200 MHz), δ : 1.28 (s br, 12H, H-3 to H-8); 1.57 (qn, 3 J = 6.7 Hz, 2H, H-2); 1.86 (qn, 3 J = 6.8 Hz, 2H, H-9); 3.41 (t. 3 J = 6.8 Hz, 2H, H-10); 3.65 (t, 3 J = 6.6 Hz, 2H, H-1); 13 C NMR (50 MHz), δ : 25.5 (C-3); 28.1 (C-8); 28.5 (C-7); 29.4 (C-4 to C-6); 32.7 (C-2, 10); 33.8 (C-9); 62.9 (C-1).

1,13-Tridecanediol 12

To a suspension of LiAlH4 (>97%, 0.93 g, 24.6 mmol, 2 eq.) in dry THF (60 ml) was added 1,11-undecanedioic acid (94%, 3 g, 12.3 mmol, 1 eq.) disolved in dry THF (150 ml) at 0°C under argon. The solution was stirred at room temperature for 2h, poured into a solution of sodium potassium tartrate at 0°C and extracted with ether. The organic layer was washed with brine, dried with MgSO₄ and concentrated *in vacuo* to give a white solid (12.47 g, 93%). TLC: (hexane-AcOEt: 7-3) Rf = 0.35; 1 H NMR (200MHz), δ : 1.28 (s br, 18H, H-3 to H-11); 1.56 (qn, 3 J = 6.6 HZ, 4H, H-2, 12); 3.64 (t, 3 J = 6.6 Hz, 4H, H-1, 13); 13 C NMR (50MHz, pyridine), δ : 26.5 (C-3, 11); 29.9 (C-4 to C-10); 33.7 (C-2, 12); 62.1 (C-1,13).

Experimental section for synthesis of 1a (n=10)

3-(Phenylsulfonyl)-cyclohexan-1-one 15

Benzenesulfinic acid sodium salt (10.25 g , 62.5 mmol , 1 eq.) was added to a solution containing cyclohexenone 14 (5 ml, 50 mmol, 1 eq.) and 30 ml of water. Then a solution of HCl 1N was added dropwise (60 ml). The reaction mixture was stirred at room temperature for 24 h and the solid formed was removed by filtration, washed with water, isopropanol and cold ether. After recrystallization in isopropanol , a white solid was obtained (5.74g, 97%)). m.p.: 83-85°C; TLC: (hexane-AcOEt: 6-4) Rf = 0,2; GC: 40°C --> 280°C (15°C/min) 11.6 min, >99%; 1 H NMR (200 MHz), δ : 1.53-1.77 (m, 2H, H-5); 2.1-2.45 (m, 4H, H-4, 6); 2.6 (d, J = 9.1Hz, 2H, H-2); 3.2-3.4 (m, 1H, H-3); 7.5-7.7 (m, 3H, H ar.-3', 4'); 7.8-7.9 (m, 2H, H ar.-2'); 13 C NMR (50 MHz), δ : 23.2 (C-5*); 23.5 (C-4*); 40.1 (C-6°); 40.2 (C-2°); 62.1 (C-3); 128.8 (C ar.-2'); 129.3 (C ar.-3'); 134 (C ar.-4'): 136.5 (C ar.-1'); 206.2 (C -1); IR (KBr): 3053 (w, =C-H); 2966, 2926 (m, C-H); 1708 (s, C=O); 1582 (w, C=C); 1450 (m); 1304 (s, SO₂); 1288 (s); 1228 (m); 1198 (m); 1159 (s, SO₂); 1138 (s); 1084 (m); 1062 (m); 912 (m); 765 (m); 728 (s, C-H ar.); 693 (m); 661, 605, 540 (m); UV (acetonitrile): λ_{max} : 222 nm (ϵ 3740), λ : 258 (ϵ 640), 264 (ϵ 905), 271 (ϵ 777); MS (EI): 238.1 (M+, 0,3); 143 (PhSO₂H₂, 0,2); 141 (PhSO₂, 0.3); 125 (PhSO, 0.4); 120,1 ((CH₂)₄SO₂, 0.4); 110 (PhSH, 0.3): 97 (M-PhSO₂, 100); 96 (cyclohexène, 17); 77 (Ph, 19); 69,1 ((CH₂)₃-CH=CH₂, 63.2); 55,1 ((CH₂-)₂-CH=CH₂, 22): microanalysis (%): calculated for C1₂H₁4O₃S (238.3) C: 60.5, H: 5.9; found C: 60.4, H: 5.7.

1,1-(Ethylenedioxy)-3-(phenylsulfonyl)-cyclohexane 16

To a solution of ketone **15** (5.3g 22.3 mmol,1eq.) in dry benzene(60ml), were added 1,2-ethanediol (0.3 ml, 1.2 eq.) and anhydrous paratoluenesulfonic acid (200 mg). The mixture was stirred under reflux for 4h, then a solution of NaHCO₃ 2M was added and the mixture was extracted with ethylacetate (3 times). The combined organic layers were washed with brine, dried with MgSO₄ and concentrated in vacuo. The solid was recrystallized in ether to obtain of white crystals (6.1g, 97%). **m.p.**: 93-95°C; **TLC**: (hexane-AcOEt: 6-4) Rf = 0.26; **GC**: 40° C --> 280° C (15° C/min): 12.8 min, >99%; ¹H NMR (200 MHz), δ : 1.3-1.6 (m. 3H, H-5, 4a); 1.61 (t, J = 12.5 Hz, 1H, H-2a); 1.65-1.73 (m, 1H, H-4e); 1.75-2.05 (m, 2H, H-6); 2.12 (ddt, J_{gem} = 12.5 Hz, ³J = 3.5 Hz, ⁴J = 2.5 Hz, 1H, H-2e); 3.21 (tt, ³J = 12.5 Hz, ³J = 3.5 Hz, 1H, H-3); 3.84-3.99 (m, 4H, O-CH₂-CH₂-O); 7.5-7.7 (m, 3H, H ar.-3', 4'); 7.8-7.9 (m, 2H, H ar.-2'); ¹³C NMR (50 MHz), δ : 21.8 (C-5); 24.5 (C-4); 33.9 (C-2*); 34.1 (C-6*); 61.3 (C-3); 64.4 (O-CH₂-CH₂-O); 107.9 (C-1); 128.9 (C ar.-2'°); 129 (C ar.-3'°); 133.6 (C ar.-4'); 136.9 (C ar.-1'); IR (KBr): 3060 (w, =C-H); 2968,2938, 2894 (m,C-H); 1583 (w, C=C); 1448 (m); 1301 (s,SO₂); 1267 (m); 1158 (s, SO₂); 1144 (s): 1082 (s); 1023 (s); 939 (m); 916 (m); 838 (m); 749 (s, C-H ar.); 718 (s); 689 (s); UV (acetonitrile): λ_{max} : 221 nm (ϵ 4970), λ : 258 (ϵ 710), 264 (ϵ 1010), 271 (ϵ 861); MS (CI, NH₃): 300.2 (MNH₄+, 100); 283.1 (MH⁺, 27); 256.1 (8): 141.1 (M-SO₂Ph, 83); **microanalysis** (%): calculated for C₁4H₁₈O₄S (282.2) C: 59.82, H: 6.32; found: C: 59.6, H: 6.4.

1,1-(Ethylenedioxy)-3-(10-hydroxydecyl)-3-(phenylsulfonyl)-cyclohexane 17

To a solution of sulfone 16 (565 mg, 2 mmol, 3eq.) and triphenylmethane (4 mg) in 5 ml of THF was added dropwise a solution of nBuLi (2 ml, 4.1 eq.) under argon at -78°C. After 10 min, the mixture was stirred at room temperature for 1 h and HMPT (1 ml) was added. It was then recooled at -78°C and a solution of bromoalcohol 9 (159 mg, 0.67 mmol, 1eq.) in 2 ml of THF was added slowly. The mixture was stirred for 2 h at -20°C and poured into a solution of saturated NH4Cl (40 ml). The solution was extracted with ether and the organic layer was washed with water and brine, dried with MgSO4 and concentrated. The residue was purified by chromatography over silica gel and eluted with hexane-AcOEt 8-2 to 6-4 to obtain a colorless oil (265 mg, 90%). TLC: (hexane-AcOEt: 6-4) Rf = 0.14; 1 H NMR (200 MHz), δ : 1.26 (s br, 14H, H-8 to H-14); 1.57 (qn br, 3 J = 6.5 Hz, 2H, H-15); 1.6-1.94 (m, 8H, H-4, 5, 6, 7); 1.98 (s, 2H, H-2); 3.64 (t, J = 6.5 Hz, 2H, H-16); 3.86-3.92 (m, 4H, O-CH₂-CH₂-O); 7.5-7.7 (m. 3H, H ar.-3', 4'); 7.81-7.87 (m, 2H, H ar.-2'); 13 C NMR (50 MHz), δ : 19 (C-5); 23.8 (C-14); 25.7 (C-7); 28.4 (C-4): 29.5 (C-10 to C-13); 30.2 (C-8); 30.5 (C-9); 32.7 (C-15); 34.5 (C-6); 35.7 (C-2); 62.9 (C-16); 63.8 et 64.8 (O-CH₂-CH₂-O); 67.4 (C-3); 108.5 (C-1); 128.7 (C ar.-3'*); 130.3 (C ar.-2'*); 133.5 (C ar.-4'); 135.8 (C ar.-1'); IR (NaCl): 3510 (m br, O-H); 3063 (w, =C-H); 2926, 2853 (s, C-H); 1585 (w, C=C); 1447 (m); 1286, 1140 (s, SO₂); 1096, 1083 (m, O-CH₂); 723, 693 (m); UV (acetonitrile): λ_{max} : 218 nm (ϵ 8600), λ : 258 (ϵ 1050), 265 (ϵ 1300), 271 (ϵ 1150); MS (IC-NH₃): 456.3 (MNH₄+, 36); 439.2 (MH+, 3.5); 299.3 (MH₂-SO₂Ph, 33); 297.2 (M-SO₂Ph, 100); 141 (SO₂Ph, 10); 98.9 (C6H₁₁O, 28).

Coupling with the other bromoalcohols

See procedure described for compound 17

Physical chemical deltails of 11 to 14 Carbon side chains

product, (n)	Formula (Yield %)	MW	Rf.*	Analysis
(11)	C ₂₅ H ₄₀ O ₅ S (97)	452	0.14	MS(CI,NH ₃): 470.3 (MNH ₄ ⁺ , 39); 453.2 (MH ⁺ , 3); 313.3 (MH ₂ -SO ₂ Ph, 41); 311.2 (M-SO ₂ Ph, 100); 141 (SO ₂ Ph, 13); 98.9 (C ₆ H ₁₁ O, 31).
				Microanalysis (%): calculated C: 66.34 H: 8.91 found C: 66.4 H: 8.7.
(12)	C ₂₆ H ₄₂ O ₅ S (96)	466	0.17	MS(CI-NH ₃): 484.2 (MNH ₄ ⁺ , 36); 467.2 (MH ⁺ , 2); 327.3 (MH ₂ -SO ₂ Ph, 27); 325.3 (M-SO ₂ Ph, 100); 256.1 (33); 141 (SO ₂ Ph, 14); 98.9 (C ₆ H ₁₁ O, 31).
(13)	C ₂₇ H ₄₄ O ₅ S (96)	480	0.17	Microanalysis (%): calculated C: 67.48 H: 9.23; found C: 67.5 H: 8.9.
(14)	C ₂₈ H ₄₆ O ₅ S (95)	494	0.20	Microanalysis (%): calculated C: 67.98 H: 9.37; found C: 67.4 H: 9.1.

^{*} TLC: hexane-AcOEt: 6-4

3-(10-Hydroxydecyl)-2-cyclohexen-1-one 18

To a solution of sulfone 17 (193 mg, 0.44 mmol) in chloroform (3 ml) and acetone (0.6 ml) was added paratoluenesulfonic acid (20 mg). The mixture was stirred at 50°C for 24 h. A solution of saturated NaHCO3 was added (10 ml) and the solution was extracted with CH₂Cl₂. The combined organic layers were washed with brine, dried with MgSO4 and concentrated in vacuo. The residue was purified on silica gel (eluting with hexane-AcOEt 8-2 to 6-4) to obtain a colorless oil (86 mg, 77%). TLC: (hexane-AcOEt : 6-4) Rf = 0.33; GC: 80 °C (1 min) --> 150 °C (15 °C/min) --> 280°C (5°C/min) 16.6 min, >99%; 1 H NMR (200 MHz), δ : 1.28 (s br, 14H, H-8 to H-14); 1.44-1.59 (m, 2H, H-15); 1.97 (qn, J = 6.4 Hz, 2H, H-5); 2.1-2.4 (m, 6H, H-4, 6, 7); 3.64 (t, J = 6.4 Hz, 2H, H-16); 5.87 (s, 1H, H-2); 13 C NMR (50 MHz), δ : 22.6 (C-5); 25.6 (C-14); 26.8 (C-4); 29.3 (C-8 to C-13); 32.6 (C-15); 37.2 (C-7); 37.9 (C-6); 62.7 (C-16); 125.5 (C-2); 166.9 (C-3); 200.1 (C-1); IR (NaCl): 3446 (m br, O-H); 3058 (w, =C-H); 2926, 2854 (s, C-H); 1665 (s, C=O); 1624 (m, C=C); 1446 (m); 1301 (m); 1152, 1125, 1078 (m, O-H); 3058 (m, =C-H); 2926, 2854 (s, C-H); 1665 (s, C=O); 1624 (m, C=C); 1446 (m); 1301 (m); 1152, 1125, 1078 (m, O-H); 3058 (m, =C-H); 2926, 2854 (s, C-H); 1665 (s, C=O); 1624 (m, C=C); 1446 (m); 1301 (m); 1152, 1125, 1078 (m, O-H); 1665 (s, C=O); 1624 (m, C=C); 1446 (m); 1301 (m); 1152, 1125, 1078 (m, O-H); 1665 (s, C=O); 1624 (m, C=C); 1446 (m); 1301 (m); 1152, 1125, 1078 (m, O-H); 1665 (s, C=O); 1624 (m, C=C); 1446 (m); 1301 (m); 1152, 1125, 1078 (m, O-H); 1665 (s, C=O); 1624 (m, C=C); 1446 (m); 1301 (m); 1152, 1125, 1078 (m, O-H); 1665 (s, C=O); 1624 (m, C=C); 1446 (m); 1301 (m); 1152, 1125, 1078 (m, O-H); 1466 (m, C=C); 1446 (m, C=C); 1446

 $\begin{array}{l} \text{CH}_2)~;~728,~693~(m)~;~\textbf{UV}~(acetonitrile):~\lambda_{max}~:~232~nm~(\epsilon~16050)~;~\textbf{MS}(EI):~252.1~(M^+,~9)~;~222.1~(8)~;~124~(12)~;~123~(C_8H_{11}O,~93)~;~110~(100)~;~97~(C_6H_9O,~65)~;~95~(C_6H_7O,~22)~;~82~(64)~;~81~(13)~;~79~(12)~;~66.9~(26)~;~55~(23)~;~\textbf{microanalysis}~(\%)~:~calculated~for~C_{16}H_{28}O_2~(252.4)~C~:~76.14,~H~:~11.18~;~found:~C~:~75.8,~H~:~10.9. \end{array}$

Elimination for the other analogues

See procedure described for product 17

Product	Formula	MW	Rf.*	Analysis
(n)	(yield %)			· ·
(11)	C ₁₇ H ₃₀ O ₂ (58)	266	0.2	m.p;.: 34-35°C GC°: Retention time: 18.4 min (purity > 99%) MS(EI): 266.1 (M ⁺ , 9); 248.1 (M-H ₂ O, 2); 236.1 (8); 124 (11); 123 (C ₈ H ₁₁ O, 83); 109.9 (100); 97 (56); 95 (C ₆ H ₇ O, 25); 82 (51); 81 (13); 79 (12); 69 (7); 66.9 (23); 55 (21). Microanalysis (%): calculated C: 76.64 H: 11.35 found C: 76.4 H: 11.6
(12)	C ₁₈ H ₃₂ O ₂ (60)	280	0.24	m.p.: 35-36°C.: GC°: Retention time: 20.1 min (purity > 99%) MS(EI): 280.3 (M+, 12); 262.1 (M-H ₂ O, 3); 250.1 (7); 150.9 (C ₁₀ H ₁₅ O, 5); 136.9 (C ₉ H ₁₃ O, 4); 124 (10); 123 (C ₈ H ₁₁ O, 84); 110 (100); 97 (51); 95 (C ₆ H ₇ O, 22); 82 (46); 81 (10); 78.9 (10); 66.9 (20); 55 (22). Microanalysis (%): calculated C: 77.07 H: 11.50 found C: 77.1 H: 11.5
(13)	C ₁₉ H ₃₄ O ₂ (93)	294	0.26	m.p.:42-43°C. : GC° : Retention time: 21.7 min (purity > 99%) MS (EI) : 294.2 (M ⁺ , 8) ; 276.1 (M-H ₂ O, 2) ; 264.1 (5) ; 151 (C ₁₀ H ₁₅ O, 5) ; 136.9 (C ₉ H ₁₃ O, 4) ; 124 (9) ; 123 (C ₈ H ₁₁ O, 77) ; 111 (8) ; 109.9 (100) ; 97 (46) ; 95 (C ₆ H ₇ O, 20) ; 82 (36) ; 81.1 (10) ; 78.9 (8) ; 66.9 (18) ; 55 (21). Microanalysis (%) : calculated C : 77.50 H : 11.64 found C : 77.4 H : 11.5
(14)	C ₂₀ H ₃₆ O ₂ (90)	308	0.28	m.p.:44-45°C. : GC° :Retention time: 23.3 min (purity > 99%) MS (EI) : 308.1 (M ⁺ , 10) ; 290.3 (M-H ₂ O, 3) ; 278.4 (6) ; 150.9 (C ₁₀ H ₁₅ O, 5) ; 137 (C ₉ H ₁₃ O, 3) ; 124 (8) ; 123 (C ₈ H ₁₁ O, 77) ; 111 (8) ; 119.9 (100) ; 97 (44) ; 95 (C ₆ H ₇ O, 19) ; 82 (30) ; 81.1 (8) ; 78.9 (7) ; 66.9 (18) ; 55 (20). Microanalysis (%) : calculated C : 77.87 H : 11.76 found C : 77.6 H : 11.4

^{*} TLC : hexane-AcOEt : 6-4

[°] GC : 80 °C (1 min) --> 150 °C (15 °C/min) --> 280 °C (5 °C/min)

Preparation of the cultures for studies of differentiation effects

The neurotrophic activities were investigated on neurons in primary culture which derived from fetal rat cerebral hemispheres (14 day-old) and the cultures were performed as described by Borg and al²⁴ with modifications. The dissociated cells were seeded at a density of 1.5×10^5 cells per 35 mm polylysine-coated Petri dish. They were then preincubated for 3h in a medium supplemented with 20% fetal calf serum, in 3 ml of a chemically defined culture medium (DMEM supplemented with insulin, transferrin, progesterone, sodium selenite and putrescine). Each compound was dissolved in ethanol and added after seeding at different concentrations ranging from 10^{-5} to 10^{-9} M. Cells were cultured without medium change for 3 days. Cultures were, then, fixed with 2% glutaraldehyde in Phosphate Buffer Saline (2.9×10^{-2} M, pH = 7.4) and stained with Coomasie Brilliant Blue. Neurons were observed and photographed under a light microscope. Control cultures were treated with ethanol only. Neurite outgrowth was quantified by the software 'NIH Image" and mean values were calculated from measuring seven photographs per tested compound.

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